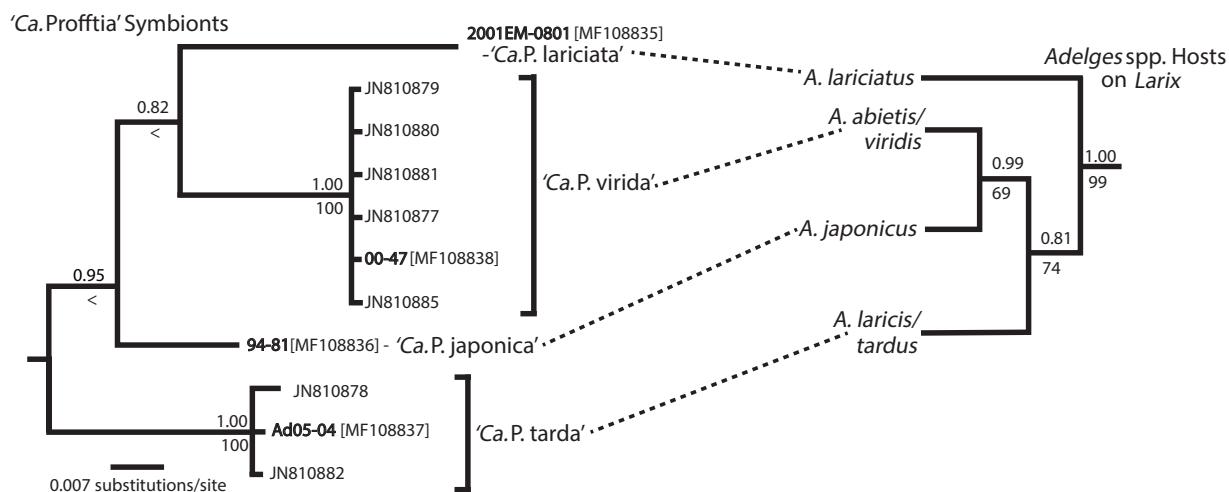
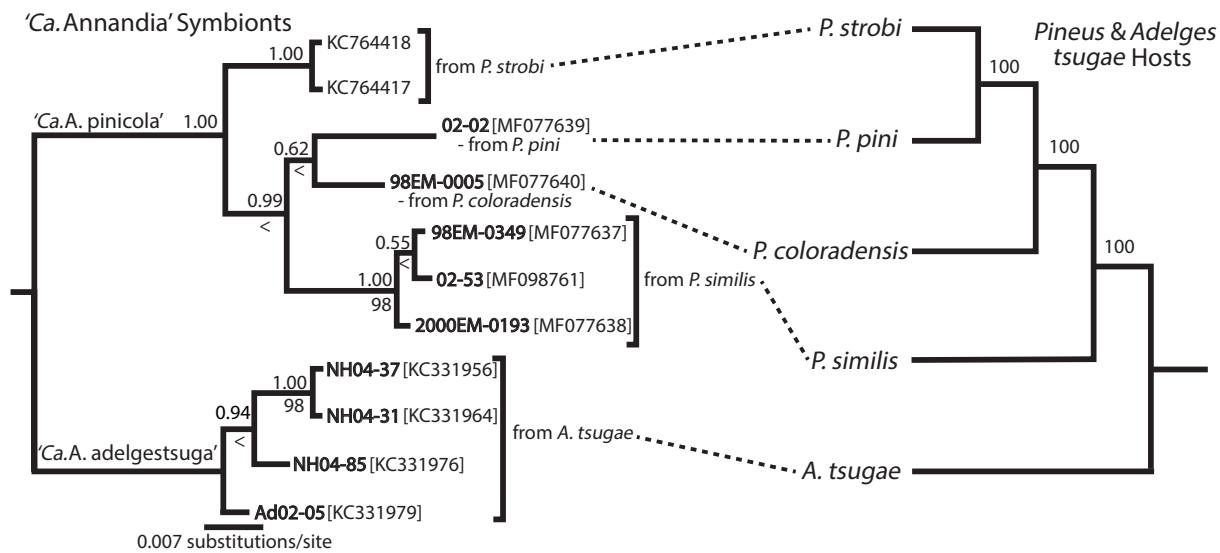


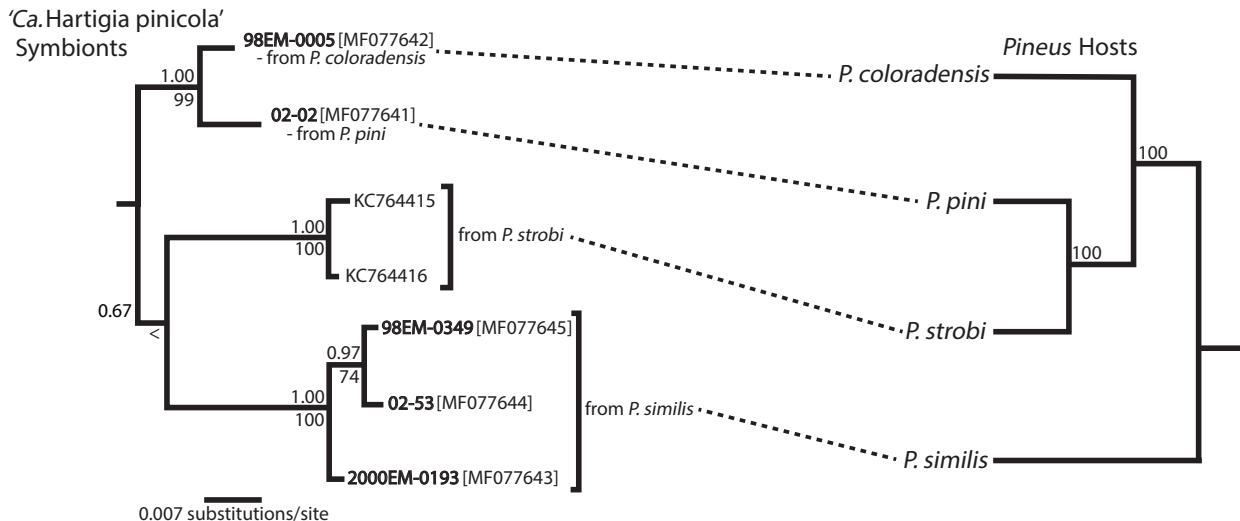
**Supplementary Figure 1.** Phylogeny of '*Ca. Vallotia*' symbionts estimated from 16S rRNA sequences (left) from five adelgid species. Adelgid phylogeny (right) was simplified from Havill *et al.* 2007. Numbers above and below branches are Bayesian posterior probabilities (BPP) and maximum-likelihood bootstrap percentages (MBP), respectively; “<” indicates BPP below 0.5 or MBP below 50%.



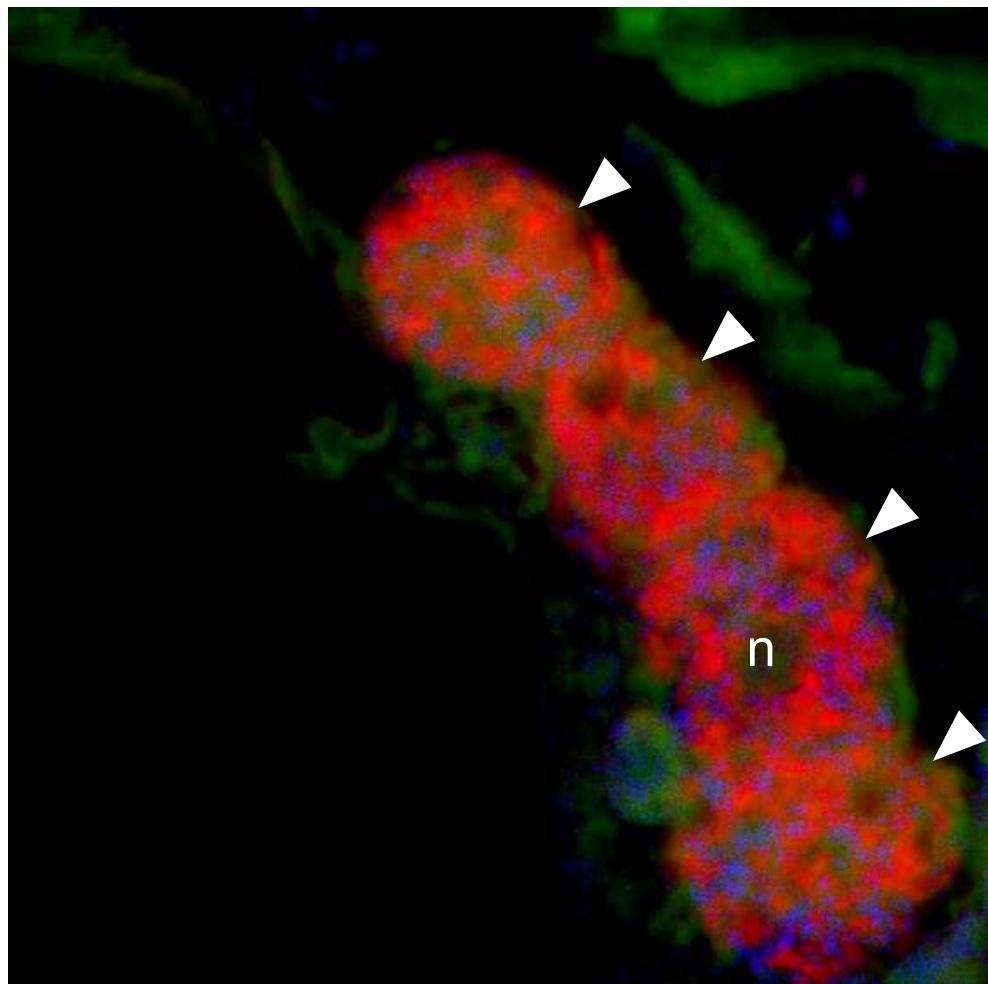
**Supplementary Figure 2.** Phylogeny of '*Ca. Proftia*' symbionts estimated from 16S rRNA sequences (left) from four adelgid species. Adelgid phylogeny (right) was simplified from Havill *et al.* 2007. Numbers above and below branches are Bayesian posterior probabilities (BPP) and maximum-likelihood bootstrap percentages (MBP), respectively; “<” indicates BPP below 0.5 or MBP below 50%.



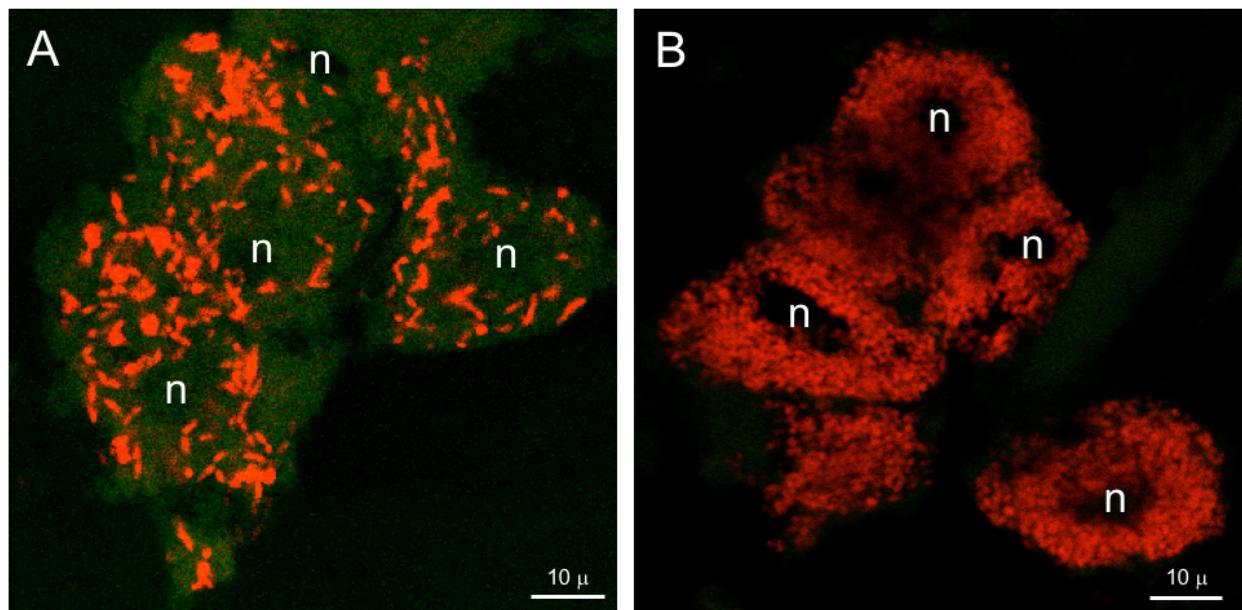
**Supplementary Figure 3.** Phylogeny of 'Ca. Annandia' symbionts estimated from 16S rRNA sequences (left) from five adelgid species. Adelgid phylogeny (right) was simplified from Havill *et al.* 2007. Numbers above and below branches are Bayesian posterior probabilities (BPP) and maximum-likelihood bootstrap percentages (MBP), respectively; "<" indicates BPP below 0.5 or MBP below 50%.



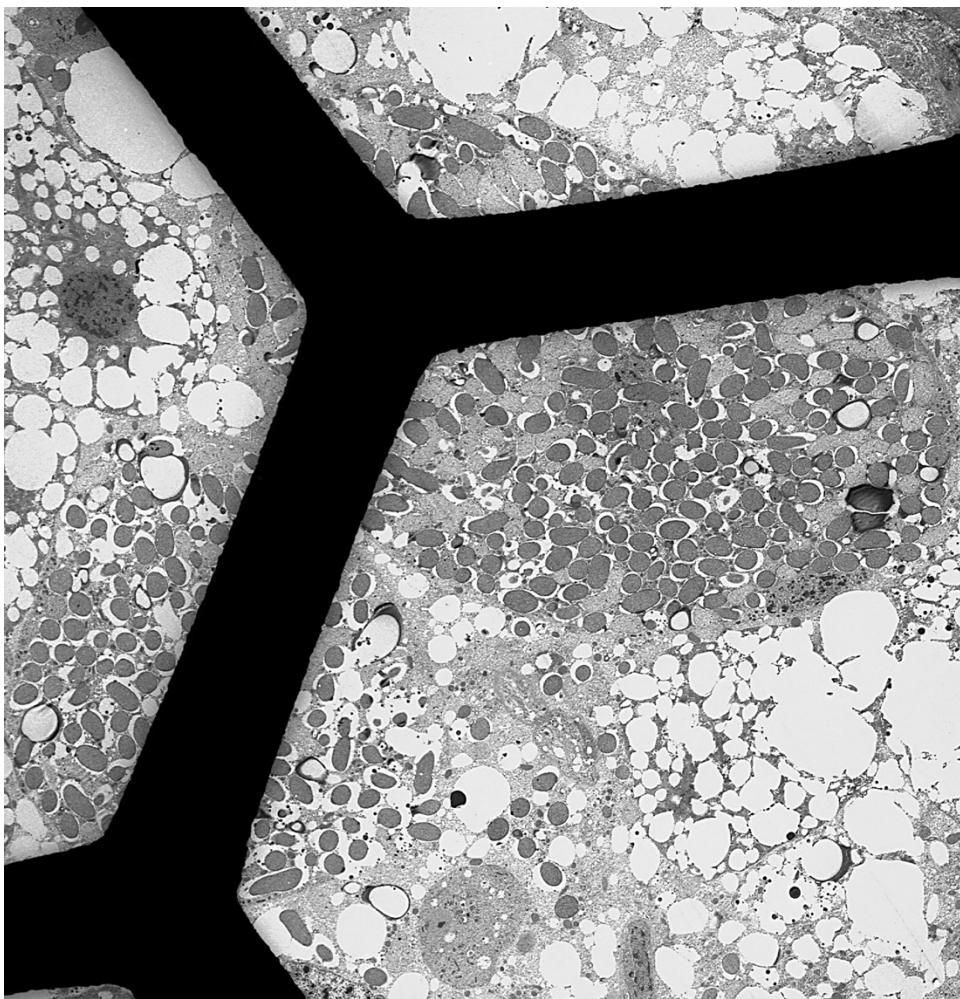
**Supplementary Figure 4.** Phylogeny of 'Ca. Hartigia' symbionts estimated from 16S rRNA sequences (left) from four adelgid species. Adelgid phylogeny (right) was simplified from Havill *et al.* 2007. Numbers above and below branches are Bayesian posterior probabilities (BPP) and maximum-likelihood bootstrap percentages (MBP), respectively; "<" indicates BPP below 0.5 or MBP below 50%.



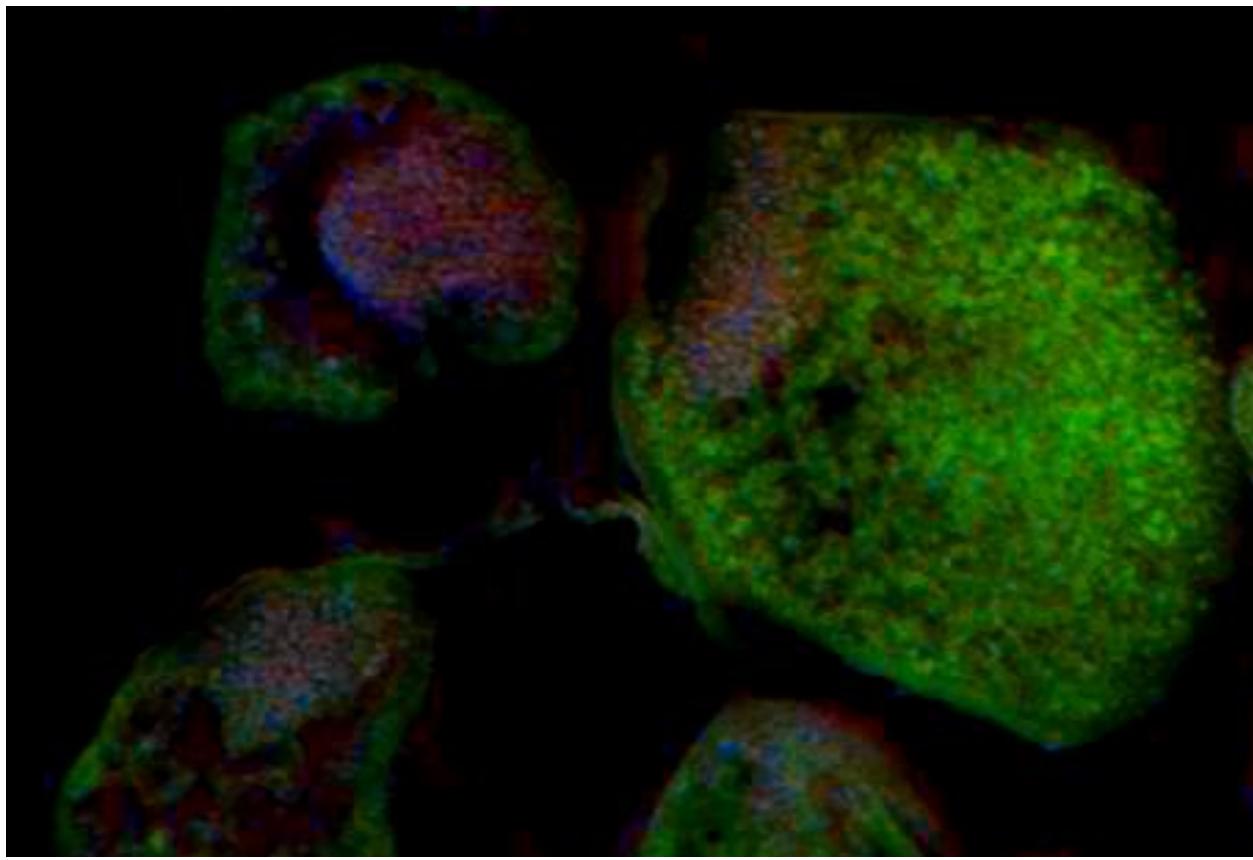
**Supplementary Figure 5.** Localization of bacteriocyte-associated endosymbionts of *Adelges laricis* by FISH. Detection of both symbionts in the bacteriomes of 4<sup>th</sup>-instar stage from larch generations (frontal section), using beta mix Al-b70 + Al-b152 + Al-b1256 labeled with Bodipy-650-14-dUTP for ‘*Ca. Vallotia tarda*’ (blue), and gamma mix Al-g1023 + Al-g1128 labeled with Alexa-568-5-dUTP for ‘*Ca. Profftia tarda*’ (red). White arrowheads point to individual bacteriocytes. Green, autofluorescence of insect cuticle; n, bacteriocyte nucleus.



**Supplementary Figure 6.** Localization of bacteriocyte-associated endosymbionts by FISH in *Adelges lariciatus* and *Adelges abietis*. **A.** FISH performed on third-instar stages of *Adelges lariciatus* to detect ‘*Ca. Vallotia*’ using specific probe b125 labeled with Alexa-568-5-dUTP (red). **B.** FISH performed on third-instar stages of *Adelges abietis* using general eubacterial probe 1507r labeled with Alexa-568-5-dUTP (red); both ‘*Ca. Vallotia*’ and ‘*Ca. Profftia*’ symbionts are localized with this general probe. Green, autofluorescence of insect cuticle; n, bacteriocyte nucleus.



**Supplementary Figure 7.** Ultrastructure of *Adelges abietis* endosymbionts residing in bacteriocytes. Ultrathin section of bacteriome in whole-mount insects, showing two distinct bacteriocytes containing two bacterial morphotypes, one light-staining and one dark-staining, corresponding to ‘*Ca. Vallotia*’ and ‘*Ca. Profftia*’ symbionts.



**Supplementary Figure 8.** Localization of bacteriocyte-associated endosymbionts in eggs from adult *P. similis* by FISH. Simultaneous detection of '*Ca. Annandia pinicola*' using GamC-440 labeled with Bodipy-650-14-dUTP (blue), and '*Ca. Hartigia pinicola*' using PinGam2-470 and PinGam2-828 labeled with Alexa-568-5-dUTP (red). Green, autofluorescence of insect cuticle.